

Bioaccumulation of some trace elements in the biota of hydrothermal fields of the Guaymas Basin (Gulf of California)

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Abstract

Data from the hydrothermally influenced Guaymas Basin of the Gulf of California are presented on the concentration and distribution of Ag, As, Au, Ba, Cd, Co, Cr, Cu, Fe, Hg, Mn, Pb, Sb, Se, and Zn in different tissues of dominant hydrothermal vent animals such as vestimentifera *Riftia pachyptila* and vesicomyid clams *Archivesica gigas* and other organisms, including *Spongia*, bivalve mollusks *Nuculana grasslei*, *Phelliactis pabista*, and crab *Munidopsis alvisca*. Chemical element content was measured by atomic absorption spectrometry (flame and graphite furnace methods) and instrumental neutron activation analysis.

In the dominant specialized taxa, the main target organs of metals were the trophosome and obturaculae of *Riftia pachyptila*, the gills and mantle of *Archivesica gigas*. The other organisms also demonstrated high bioaccumulation of metals. Especially high levels of most of the metals (excluding Mn) were detected in the soft body of *Nuculana grasslei*. The highest Mn content was found in the whole body of *Spongia*. Bioconcentration factor of the trace metals studied varies within three orders of magnitude from 5 (Mn) to 3•10⁴ (Cd). This testifies apparently a selectivity of trace metal bioaccumulation by the organisms which is determined by metal bioavailability independently of metal concentration in the water column. Variability in the molar ratio Fe/Mn allows us to assume that these metals undergo fractionation during migration from the hydrothermal fluids to the interior organs of animals. Insignificant differences between the Cd, Cu, Fe, Hg, Pb, and Zn levels in the Guaymas Basin vent clams versus that in the bivalve mollusks from polluted areas of the Gulf of California might suggest that the metal bioavailability play an important role in the bioaccumulation.

Key words: trace metals; bioaccumulation; Guaymas Basin; hydrothermal communities.

Resumen

Se presentan los datos sobre la concentración y distribución de Ag, As, Au, Ba, Cd, Co, Cr, Cu, Fe, Hg, Mn, Pb, Sb, Se, and Zn en diferentes tejidos de los animales predominantes en las ventilas hidrotermales tales como la vestimentifera <u>Riftia pachyptila</u>, la almeja vesicomyidae <u>Archivesica gigas</u>, así como otros organismos, que incluyen <u>Spongia</u>, moluscos bivalvos <u>Nuculana grasslei</u>, <u>Phelliactis pabista</u>, y el cangrejo <u>Munidopsis alvisca</u>. El contenido de los elementos químicos fue medido por medio de espectrofotometría de absorción atómica (métodos de flama y de horno de grafito) y por el análisis instrumental de la activación neutrónica.

En la taxa especializada dominante, los órganos principales en los cuales se acumularon los metales fueron el trofosoma y el obturaculae de <u>Riftia pachyptila</u>, las branquias y el manto de <u>Archivesica gigas</u>. Los otros organismos también mostraron alta bioacumulación de metales. Especialmente altos niveles de la mayoría de los metales (excepto Mn) fueron detectados en el cuerpo suave de <u>Nuculana grassley</u>. El contenido más alto de Mn fue encontrado en todo el cuerpo de <u>Spongia</u>. El factor de bioconcentración para los metales traza estudiados varía en tres ordenes de magnitud desde 5 (Mn) a $3 \cdot 10^4$ (Cd). Esto aparentemente evidencia cierta selectividad en la bioacumulación de los metales traza por los organismos, la cual es determinada por la biodisponibilidad independientemente de la concentración de metales en la columna del agua. La variabilidad de la razón molar de Fe/Mn nos permite asumir que estos

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metales durante la migración desde los fluidos hidrotermales hacia los órganos internos de animales están sujetos a fraccionación. Las diferencias insignificantes entre los niveles de Cd, Cu, Fe, Hg, Pb y Zn en las almejas de las ventilas de la Cuenca de Guaymas y los niveles de los moluscos bivalvos de áreas contaminadas del Golfo California podrían sugerir que la biodisponibilidad de los metales juega un papel importante en la bioacumulación.

Palabras clave: metales pesados; bioacumulación; Cuenca Guaymas; comunidades hidrotermales.

1. Introduction

Deep-sea hydrothermal vent fields are inhabited by many thermophilic organisms that are able to survive under extreme physical and chemical conditions, including high loads of trace metals and redox gases (Anonymous, 1995; Sarradin et al., 2008; Van Dover, 2000). An exceptional feature of the semi-enclosed Guaymas Basin (Gulf of California) hydrothermal vent field is the thick organic-rich sedimentary cover on the seafloor. This cover is a result of high sedimentation rates due to Colorado River sediment input directly before dam construction or tidal resuspension of previously supplied terrigenous sediments in the Upper Gulf of California (Calvert, 1966; Carriquiry and Sánchez, 1999; Carriquiry et al., 2001) and biogenic particles from the highly productive euphotic zone (Lonsdale *et al.*, 1980; De la Lanza-Espino and Soto, 1999; Thunell, 1998). High-temperature fluids are discharged to the surrounding seawater through the vents and by ascending through the overlying sediments, which are rich in Mn. This leads to the enrichment of fluids for Mn relative to Fe (Von Damm et al., 1985), which is a characteristic feature of Guaymas Basin fluids compared to other vent fields. From this standpoint, it is interesting to examine the trace metal and other element contents of the hydrothermal organisms inhabiting the vent environments, especially since the relationships between organisms from the Guaymas Basin and metals have not been thoroughly studied thus far. There are few articles devoted to the chemical composition of Guaymas Basin organisms, and those that do exist focus on heavy metals in dominant taxa inhabiting vent environment, such as vestimentifera Riftia (Lein et al., 1989; Lukashin et al., 1990; Ruelas-Inzunza et al., 2005), bivalve mollusks Archivesica gigas (Lein et al., 1989; Lukashin et al., 1990) and Vesicomya gigas (Ruelas-Inzunza et al., 2003). Meanwhile, little is known about metal bioaccumulation in other taxa that live in the vent areas and their periphery but are not dependent upon symbiotrophy. The dominant hydrothermal fauna studied here is represented by the highly specialized taxa vestimentifera Riftia pachyptila and clam Vesicomyidae Archivesica gigas, both of which are nutritionally dependent on the chemosynthetic bacterial community. We also consider other organisms such as Spongia, bivalve mollusk Nuculana grasslei, Phelliactis pabista, and crab Munidopsis alvisca.

The aim of this work was to study features of the

distribution of 15 chemical elements (including Ag, As, Sb and Au, as well as organic carbon or Correl, for which there were no published data on these organisms) in the fauna microhabitats, the tissues and whole bodies of the hydrothermal organisms from the Guaymas Basin, including the dominant specialized symbiotrophic and the peripheric taxa. Besides, it is interesting to estimate the concentration factors of metals in organisms relative to the water column. There are a some mining complexes located in the Baja California peninsula, as well as other different types of industrial and urban activities along the coastal zone of the Gulf of California, which sometimes could produce a deficiency of oxygen, acidic conditions and high discharges of heavy metals into the ecosystems. We aim to determine whether there are any differences in the heavy metal content of the organisms exposed to hydrothermal and anthropogenic impacts.

2. Materials and methods

2.1. Geological setting and faunal distribution

The Southern trough of the Guaymas Basin's hydrothermal field is located at 2020 m depth (27°00.70'N, 111°24.40' W), presented in Figure 1. The intense tectonic activity in this area is caused by the displacement of the Baja California peninsula towards the NW at a spreading rate of about 6 cm per year (Klitgord et al., 1974; Lonsdale et al., 1980). An exceptional feature of the semi-enclosed Guaymas Basin's hydrothermal vent field is the thick organic-rich sedimentary cover of the seafloor. This is a result of high sedimentation rates (1-2 mm per year) due to significant terrigenous input from the Colorado River (Calvert, 1966) before its complete damming in 1956, tidal resuspension of sediments from the Colorado River delta in the Upper Gulf of California, and the high productivity of the euphotic zone of the Gulf of California (De la Lanza-Espino and Soto, 1999; Thunell, 1998). The surface sediments of the Guaymas Basin also have a Mn-oxide-rich and (relatively) Fe-oxide-rich turbidite layer that affects the distribution of C, Fe, Mn, S and some trace metals (Otero et al., 2003). Iron is mainly pyritized in the sediments, while Mn is found predominantly in carbonates (41±12 %) and is associated with pyrite to a much lesser degree; Co, Cr, Cu, Ni and Zn were highly pyritized (> 80 %) in



Figure 1. Study area and location of sampling sites in the Guaymas Basin's hydrothermal field of the Gulf of California

the sediments of the Guaymas Basin (Otero et al., 2003). The low-temperature hydrothermal mineral associations on the floor of the Guaymas Basin are represented by opal and barite, while pyrrhotite, sphalerite and chalcopyrite are the dominant ore minerals in high-temperature areas (Bogdanov et al., 2004). It is interesting to note that both mineral formations contain oil hydrocarbons, with a content of C_{org} in the surface sediments ranging from 0.15 % (high-temperature area) to 2.23 % (low-temperature area) (Bogdanov et al., 2004) and reaching up to 6.21% in some deposits saturated with hydrocarbons (Peresypkin, personal communication). More than a hundred high-temperature hydrothermal mounds (black smokers) in an area of 30 km² were discovered and described by Lonsdale *et al.* (1980). Sulfide chimneys commonly grow through overlying sediments and can reach heights of more than 25 m. Hightemperature fluids (maximum temperatures of up to 315°C) are emitted from the vents into the surrounding water. Warm fluids flow through the chimney walls and ascend past the sedimentary cover, which is enriched with organic matter. This leads to a complicated transformation of organic matter into hydrocarbons and methane (Von Damm et al., 1985) that is characteristic of the Guaymas Basin fluids compared to other known vent fields.

Of the variety of factors determining fauna distribution, temperature and substratum characteristics are the easiest to estimate during bottom observation. Precise data is lacking in most cases, but numerous visual observations accompanied by temperature estimations have found that black smokers have a temperature in the range 275-400°C, while white smoker temperature varies from 100 to 250°C (BRIDGE Workshop Report, 1994). The heat extreme for most vent zone inhabitants seems to be about 25-40°C. A few prominent zones can be identified in the Guaymas Basin's hydrothermal vent field, each of which is dominated by certain megafaunal groups: 1) the euthermal or shimmering water zone (ambient temperature of about 25-30°C), where vestimentiferans commonly live; 2) the oligothermal zone (temperatures of 3-6 to 25°C), which is populated by vesicomyid clams and mytilid mussels; 3) the periphery of the vent zone (near-field, with very low or absent temperature anomalies), where specialized suspension-feeders consuming bacteria are the predominant taxa; and 4) the periphery of the vent zone (far-field, without temperature anomalies), which is occupied by non-vent suspension-feeders (Galkin, 2002).

Abundant settlements of vestimentifera *Riftia pachyptila*, reaching up to 1 m length and occupying areas of up to

hundreds m², were detected in the shimmering water at the hydrothermal chimney surfaces. The basic group of fauna inhabiting the soft sedimentary cover is the vesicomyid clam Archivesica gigas whose settlements can accumulate up to hundreds specimens per m². These communities of organisms are nutritionally dependent on the chemosynthetic bacterial community and are typically surrounded by accumulations of the bivalve mollusks Nuculana grasslei (Allen, 1993). Sediments soaked with hydrocarbons serve as a substratum for these organisms. Chimney walls and bases are inhabited by Munidopsis alvisca crabs (predator), Spongia (filter-feeder) and Phelliactis pabista (filter-feeder and predator). The latter were often attached to the shells of the vesicomyid clam Archivesica gigas. Thick bacterial mats (with a thickness of up to a few cm) cover significant areas (hundreds of m²).

2.2. Fluid and animal collection

The material studied involved water samples taken from the habitat of the vestimentifera Riftia (two shimmering water samples from St. 4700, at 27°00'48 N, 111°24'74 W) as well as from the habitat of the vesicomyid clam Archivesica gigas (two water samples from the low temperature diffuser habitats St.4709 (26°00'34 N, 111°24'78 W). Twenty-five samples of different organs and tissues from the dominant bottom organisms were also collected in the Guaymas Basin vent field (Figure 1). These samples were taken during the 49th cruise of the Russian research vessel "Akademik Mstislav Keldysh" (16-20 October, 2003) using the "Mir-1" and "Mir-2" manned submersibles. Water samples were collected by the "Mir" submersibles using 700 ml titanium syringes designed for sampling of hot fluids. Aboard the ship, the samples were immediately filtered trough a 1 µm pore size nucleopore filter and placed afterwards into acidwashed high density polyethylene bottles and acidified to pH 2 with nitric acid (super pure MERCK). Water samples were stored in the refrigerator until analysis in the stationary laboratory, which determined the content of total dissolved chemical elements.

The samples of bottom fauna were collected using the "slurp-gun" and sieve nets operated by the submersibles. Each sample was washed with deionized water aboard the ship to eliminate sea salts and then measured and freshly dissected into the main organs; some individuals were taken as a whole body. The shells of mollusks and the vestimentiferan tubes were rinsed with deionized water after being separated from the interior organs, but they were not treated with a 10% solution of HNO₂ (as was done by Kádár and Costa, 2006), as this could have eliminated bacterial overgrowth and extracted the adsorbed fraction of metals from the surfaces of these organs. This method of shell preparation allowed us to study the total amount of metals in the external organs of invertebrates, whether incorporated into the shell or adsorbed onto their surface. The samples were dried at 60°C and stored in insulated plastic bags until analysis.

2.3. Pretreatment and chemical analysis of samples

The animal samples (ranging from 10 to 50 mg in weight) were carefully powdered and digested with a 1 ml mixture (2:1) of ultra pure Merck concentrated nitric acid (69% v/v) and 30% hydrogen peroxide in Teflon vessels with MWS-2 microwave system (Berghof, Germany). Before analytical measurement, the water samples of fluids and the obtained solutions of digested tissues were diluted to 1:5 and 1:10, respectively, using high purity deionized water. The concentrations of the chemical elements in fluids were determined by atomic absorption spectrophotometry using a KVANT-2A instrument (flame version) and a KVANT-ZETA instrument (graphite furnace version) at the P.P. Shirshov Institute of Oceanology (Moscow, Russia) of the Russian Academy of Sciences. Some elements were determined by instrumental neutron activation analysis at the V.I. Vernadsky Institute of Geochemistry and Analytical Chemistry (Moscow, Russia) of the Russian Academy of Sciences. The details of the analytical methods employed are given in Table 1. The detection limits for flameless AAS measurements were 0.5 μ g l⁻¹ (Ag), 0.7 μ g l⁻¹ (As), 0.1 μg l⁻¹ (Cd and Cu), 0.2 μg l⁻¹ (Co and Cr), 0.1 μg l⁻¹ (Fe), 0.2 μ g l⁻¹ (Mn), 1 μ g l⁻¹ (Sb and Se) and 0.05 μ g l⁻¹ (Zn). Standard solutions for metal determination were prepared using certified State Standard Samples (GSO). Accuracy of the analysis was controlled through the use of international reference materials: NIST SRM 2976 (mussel tissue), IAEA MA-A-2/T (fish flash) and GSD-7. A comparison of the metal concentrations of certified standard reference materials with the measured values revealed a percentage recovery rate of between 92-95% for Cu, Fe, Mn and Zn; 86-90% for Ag, Cd, Co, Cr and Pb; 82-85% for Hg, Sb, and Se. Determination of organic carbon was carried out in 14 samples using a Carbon Express-Analyzer AN-2975 M; with an accuracy of 0.1 %.

Tabla 1. Analytical techniques and precision of the methods

Elements, object	Analytical technique	Precision %
Cu, Fe, Mg and Zn in fluids	Flame Atomic Absorption Spec- trophotometry (AAS)	5
Ag, As, Cd, Co, Cr, Pb, and Sb in fluids	Graphite furnace AAS	12
As, Au, Ba, Co, Cr, Fe, Sb, Se in organisms	Instrumental neutron activation analysis	15
Cu, Mn and Zn in organisms	Flame Atomic Absorption Spec- trophotometry (AAS)	8
Ag, Cd, and Pb in organisms	Graphite furnace AAS	10
Hg	Cold vapor concentration - Graph- ite furnace AAS after reduction with SnCl,	14

2.4. Data treatment

Standard statistical software was used for the data treatment. The bioconcentration factor (F_c) was also calculated accordingly to a relation:

$$F_c = C_m / C_{water}$$

where C_m – metal content in the whole organism (mg kg⁻¹ dry weight), calculated from the concentrations of each metal in organs, constitutioning whole body of the animal, on the base of the percentage of mass of each organ, and C_{water} is the concentration of metal in corresponding hydrothermal solution (mg kg⁻¹).

3. Results

3.1. Heavy metals in the fauna microhabitats

The results of element determination for the four fluid samples are listed in Table 2. Samples collected from shimmering water ("hot fluids" of the first type) at St. 4700 had a weakly acidic character (pH = 5.4), while samples collected from a nearby diffuser vent ("warm fluids" of the second type) at St.4709 were more alkaline (pH = 7.49). In both cases, pH is rather low in comparison to ocean water (usually pH 8.0-8.2; Millero, 1996). Fluids of the first type are more deficient in Mg than fluids of the second type. The Mg concentrations of both types of fluids are somewhat lower than observed in ocean water. In both cases, the trace metal contents are much higher in the fluid samples relative to the reference ocean water. According to our data, fluids of the first type are enriched in As, Cr, Cu, Fe, Mn, and Pb relative to fluids of the second type, but deficient in Ag, Cd, Sb and Zn. The Mn and Zn concentrations show the largest difference between the two types of fluids (Table 2).

3.2. Trace metals in the Guaymas Basin organisms

Average values for metal content (mg kg⁻¹ dry weight) in the different tissues of organisms and, in some cases, the whole bodies, are presented in Table 3. The highest metal contents over all the samples studied were found for nutritionally important Ba, Fe, Mn and Zn (with peak concentrations of up to thousands of mg kg⁻¹), followed by Cu (up to hundreds of mg kg⁻¹) and Ag, As, Cr, Pb and Sb (up to dozens of mg kg⁻¹). Contents of Cd, Co, Hg and Se did not exceed 5 mg kg⁻¹, while Au had the lowest content, typically less than 0.1 mg kg⁻¹. The differences in the maximal and minimal contents of the pooled metals reached up to six orders of magnitude.

Vestimentifera Riftia is a highly specialized symbiotrophic and the most abundant organism the East Pacific Rise. It inhabits shimmering waters and is nutritionally dependent on the reduced substances (mainly H₂S) contained in the hydrothermal fluids. Chemical analysis of separate organs of mature individuals (which have a tube length of up to 51 cm) have revealed two groups of elements that can be distinguished based on their contents: i) As, Ba, Cu, Fe, Mn and Zn with average contents between 10 and 450 mg kg⁻¹ (Figure 2a); ii) Ag, Cd, Co, Cr, Hg, Pb, Sb, and Se with average contents below 10 mg kg⁻¹ (Figure 2b). Different organs of Riftia show various capacities for accumulating metals. The differences are obviously associated with distinctions in their functions. Ag, Co, Fe and Hg are accumulated to a greater extent in trophosomes relative to the other organs. The highest Ba concentration was found in Riftia's tube, which consists primarily of chitin and is covered with a thick layer of bacterial biomass. The highest As, Cd, Mn, Se and Zn contents were found in the obturaculae, which is the anterior lociniate (or wing) end directed toward fluids. The opisthosome is the posterior end and is attached to the substratum. Relative to other Riftia's organs it is noticeably enriched in Co, Cu and Sb. The vestimentum, which represents the middle muscular, collar-like part of the body, is enriched in Pb relative to other organs. Gonads, which are responsible for reproduction, accumulate more Cr than other organs (Figure 2b). The highest organic carbon ($C_{_{OTR}}$) content (28.53 %) was detected in the trophosome. This occurs quite naturally due to accumulation of endosymbiotic bacteria. Meanwhile, the lowest Corr value (20.32 %) was found in the vestimentifera tube.

The second dominant specialized taxon is the vesicomyid clam *Archivesica gigas*, whose gills are the main target organ of Zn (3110 mg kg⁻¹) and Cu (42.5 mg kg⁻¹), along with very similar value of 45.54 mg kg⁻¹ in mantle, Cr (21.4 mg kg⁻¹), Co (1.3 mg kg⁻¹), and Se (1.5 mg kg⁻¹). The content of these metals in gills is much higher (up to one

Table 2. Concentrations of elements in the water samples from the Guaymas Basin's hydrothermal field (average ±standard deviation)

Water sample type	Ag, nM	As, nM	Cd, nM	Co, nM	Cr, nM	Cu, µM	Fe, μM	Mg, mM	Mn, μM	Pb, nM	Sb, nM	Zn, μM
First type (n=2): hot, pH 5.40	2±1	700±80	2±1	190±130	760±230	0.66±0.16	20.6±2.5	34.00±1.2	43.6±3.6	41±5	30±11	0.32±0.11
Second type (n=2): warm, pH 7.49	26±8	100±50	4±1	160±110	470±120	0.29±0.07	7.5±1.4	43.10±2.32	1.48±0.06	16±9	50±13	18.75±1.24
Ocean water (Li, 1991)	0.02	22	0.7	0.03	5	0.003	0.004	52.2	0.001	0.01	1.2	0.011

specimens)	Tissue	Ag	As	Au		Ba	Cd	Co	Cr	Cu
	trophosome	3.46±1.2	7.12±1.:	5 0.018±0	0.004	34±3	1.86±0.32	3.24±1.53	1.1±0.4	15.4±2.6
	obturaculae	0.62±0.23	6.3±1.8	0.019±0	0.005 1	1375±220	12.1±1.7	0.35±0.27	0.69±0.2	7.9±2.7
<i>Riftia pachyptila</i> (n=3),	opisthosome	1.25±0.15	0.73±0.	1 0.023±0	0.007	56±5	2.44±0.32	0.97±0.73	2.87±1.45	0.6±0.2
tube length= 10-21.5 cm	vestimentum	0.43±0.12	3.67±0.9	0.03±0	0.01	290±45	2.0±0.4	0.32±0.15	1.47±0.43	7.9±3.2
	tube	nd	3.59±1.0	0 0.019±0	0.007	335±38	4.04±0.42	1.78±0.64	5.44±1.38	11.3±3.5
	trophosome	0.92	17.8	0.00	7	7	0.44	0.34	3.14	34.0
	obturaculae	0.74	46.4	0.02	5	42	2.02	0.28	2.6	24.3
Riftia pachyptila (n=1)	opisthosome	0.62	10.1	nd		38	0.60	2.95	0.46	42.8
tube length= 51 cm	vestimentum	0.4	2.25	0.01	4	45	0.08	0.37	0.41	8.7
	gonades	0.48	0.34	0.00	7	115	0.07	1.28	3.76	4.6
	tube	nd	1.93	nd		200	0.38	0.48	4.78	26.3
	tube, posterior end	0.05	1.22	0.00	7	70	1.06	0.3	3.78	4.8
<i>Riftia pachyptila</i> (n=1),	tube, middle part	nd	6.25	0.03	7	67	0.58	0.53	0.59	21.3
tube length= 115 cm	tube, anterior end	nd	28.8	0.00	1	185	0.44	0.36	0.78	8.6
	whole soft tissues	64.0±7.3	56.3±18.	.2 0.034±0	.013 1	1440±208	4.0±0.35	0.77±0.25	3.9±2.0	879±75
Nuculana grasslei (n=15)	shell	1.24±0.16	2.33±0.7	1 0.003±0	.002	58±39	0.34±0.05	0.23±0.1	2.65±0.7	16.6±4.8
	foot	0.37	3.57	0.01	8	73	3.62	0.41	5.57	9.2
Archivesica gigas (n=1),	gills	1.22	1.01	0.02	4	280	1.12	1.29	21.4	42.5
shell lengui-/5 mm	mantle	7.29	4.14	0.08	5	725	4.32	0.29	4.03	45.5
Archivesica gigas (n=1), shell length=88 mm	shell	0.03	1.16	0.00	3	870	3.64	0.27	0.61	8.5
Phelliactis pahista (n=1)	whole	nd	8.15	nd		140	5.70	0.28	1.29	69.7
()	chitin	0.81	1.92	0.00	3	21	2.86	0.3	0.25	51.7
Munidopsis alvisca (n=1)	spawn	0.44	2.65	nd		47	0.34	0.18	0.65	39.9
······································	remaining	1.43	3.92	0.02	2	125	4.32	0.36	1.72	122
Spongia (n=1)	whole	0.91	9.5	0.01	2	225	6.64	0.97	1.67	80.7
Taxon (number of specimens)	Tissue	Cu	Fe	Hg	Mn	Pb	Sb	Se	Zn	C _{org} , %
Taxon (number of specimens)	Tissue trophosome	Cu 15.4±2.6	Fe 785±80	Hg 1.66±0.3	Mn 10.2±3	Pb 3.8 0.14±0	Sb 0.08 4.95±	Se 1.2 0.41±	Zn 0.1 255±27	C _{org} , % 24.2±2.3
Taxon (number of specimens)	Tissue trophosome obturaculae	Cu 15.4±2.6 7.9±2.7	Fe 785±80 92±8	Hg 1.66±0.3 1.21±0.42	Mn 10.2±3 19.3±7	Pb 3.8 0.14±0 7.5 24.6±1	Sb 0.08 4.95± 11.3 8.19±	Se 1.2 0.41± 2.0 0.93±(Zn 0.1 255±27 0.23 231±27	C _{org} , % 24.2±2.3 nd
Taxon (number of specimens) <i>Riftia pachyptila</i> (n=3), tube length = 10,21,5 cm	Tissue trophosome obturaculae opisthosome	Cu 15.4±2.6 7.9±2.7 0.6±0.2	Fe 785±80 92±8 nd	Hg 1.66±0.3 1.21±0.42 4.34±1.35	Mn 10.2±3 19.3±7 10.1±2	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0	Se 1.2 0.41± 2.0 0.93±0 0.25 1.31±0	Zn 0.1 255±27 0.23 231±27 0.25 410±46	C _{org} , % 24.2±2.3 nd nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm	Tissue trophosome obturaculae opisthosome vestimentum	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2	Fe 785±80 92±8 nd 10±3	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37	Mn 10.2±3 19.3±7 10.1±2 15.5±5	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±0	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.1 5.67±	Se 1.2 0.41± 2.0 0.93±(0.25 1.31±(2.1 0.34±(Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12	C _{org} , % 24.2±2.3 nd nd 22.9±1.4
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm	Tissue trophosome obturaculae opisthosome vestimentum tube	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5	Fe 785±80 92±8 nd 10±3 102±14	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.1 5.67± 0.93 1.44±	$\begin{array}{ccc} & & & \\ & 1.2 & 0.41 \pm \\ 2.0 & 0.93 \pm (\\ 0.25 & 1.31 \pm (\\ 2.1 & 0.34 \pm (\\ 0.2 & 0.26 \pm \end{array} \end{array}$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0	Fe 785±80 92±8 nd 10±3 102±14 463	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 1.10	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.33	Se 1.2 0.41± 2.0 0.93±(0.25 1.31±(2.1 0.34±(0.2 0.26± 3 1.84	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101	C _{org} , % 24.2±2.3 nd 22.9±1.4 20.3±1.7 28.5
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3	Fe 785±80 92±8 nd 10±3 102±14 463 451	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 1.10 0.05	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.33 5 11.3	Se 1.2 0.41± 2.0 0.93±(0.25 1.31±(0.2 0.26± 3 1.84 3 4.33	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1),	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8	Fe 785±80 92±8 nd 10±3 102±14 463 451 131	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47± 2.5 3.65±0 1.10 0.02 3.8 0.03	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.13 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7	Se 1.2 0.41± 2.0 0.93±(2.25 1.31±(2.1 0.34±(0.2 0.26± 3 1.84 3 4.33 7 1.5	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47± 2.5 3.65±0 1.10 0.02 3 0.02 3 0.02	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44±0 0 6.33 5 11.3 5 13.7 5 13.7 0 2.09	Se 1.2 0.41± 2.0 0.93±(0.25 1.31±(0.2 0.26± 3 1.8* 4.3: 7 7 1.5 0 0.3*	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4
Taxon (number of specimens) <i>Riftia pachyptila</i> (n=3), tube length= 10-21.5 cm <i>Riftia pachyptila</i> (n=1), tube length= 51 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47± 2.5 3.65±0 1.10 0.05 3.00 0.04	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72	Se 1.2 0.41± 2.0 0.93±(2.25 1.31±(2.1 0.34±(0.2 0.26± 3 1.84 3 4.33 7 1.5 9 0.37 2 1.53	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 1.10 5 0.05 3 0.05 3.00 0.47 0.75	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 5 13.7 0 2.09 7 1.72 5 0.74	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.25 $1.31\pm$ (2.1 $0.34\pm$ (0.2 $0.26\pm$ 3 1.84 3 4.33 7 1.55 2 1.53 4 1.6	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 4 4	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 1.10 5 0.05 3 0.05 3.00 0.43 0.75 0.75	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72 5 0.74 3 0.21	$\begin{array}{c cccc} & & & & & \\ \hline 1.2 & 0.41 \pm \\ 2.0 & 0.93 \pm (\\ 2.25 & 1.31 \pm (\\ 2.1 & 0.34 \pm (\\ 0.2 & 0.26 \pm \\ 3 & 1.84 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 0 & 0.37 \\ 2 & 1.53 \\ 4 & 1.6 \\ \hline 1 & 0.42 \end{array}$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 5 44 2 40	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 1.10 5 0.05 3 0.05 3.00 0.44 0.75 0.75 2.45 0.75 2.45 0.75	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72 5 0.21 5 0.21 5 0.21 5 0.21 5 0.52	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 4 175 7 82 5 173 6 44 2 40 9 73	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end	$\begin{array}{c} Cu\\ 15.4 \pm 2.6\\ 7.9 \pm 2.7\\ 0.6 \pm 0.2\\ 7.9 \pm 3.2\\ 11.3 \pm 3.5\\ 34.0\\ 24.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 4.8\\ 21.3\\ 8.6\end{array}$	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 5.4 0.47±1 5.5 0.03 3.00 0.47 0.72 0.72 0.72 2.44 3.68 3.68	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.11 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72 5 0.74 3 0.21 5 0.52 8 4.4	$\begin{array}{c ccccc} & & & & & \\ \hline 1.2 & 0.41 \pm \\ 2.0 & 0.93 \pm (\\ 0.25 & 1.31 \pm (\\ 2.1 & 0.34 \pm (\\ 0.2 & 0.26 \pm \\ 3 & 1.84 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 1.5 \\ 2 & 1.53 \\ 4 & 1.6 \\ 1 & 0.43 \\ 2 & 3.79 \\ 1.23 \end{array}$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 7 82 5 173 6 44 2 40 9 73 5 42	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3	Pb 3.8 0.14±0 7.5 24.6±1 2.4 1.52±0 5.4 0.47±1 2.5 3.65±0 5.4 0.47±1 5.5 0.03 3.00 0.47 0.73 0.73 2.44 3.68 3.2 24.3±1	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.17 5.67± 0.93 1.44± 0 6.32 5 11.3 5 13.7 5 0.74 3 0.21 5 0.72 5 0.52 8 4.4 0.02 13.1±	$\begin{array}{c ccccc} & & & & & \\ \hline 1.2 & 0.41 \pm \\ 2.0 & 0.93 \pm (\\ 2.25 & 1.31 \pm (\\ 2.1 & 0.34 \pm (\\ 0.2 & 0.26 \pm \\ 3 & 1.84 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 1.5 \\ 4 & 1.6 \\ 1 & 0.42 \\ 2 & 3.74 \\ 1.22 \\ 2.8 & 7.3 \pm 2 \end{array}$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 6 44 2 40 9 73 5 42 2.1 580±60	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd
Taxon (number of specimens)Riftia pachyptila (n=3), tube length= 10-21.5 cmRiftia pachyptila (n=1), tube length= 51 cmRiftia pachyptila (n=1), tube length= 115 cmNuculana grasslei (n=15)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23	$\begin{array}{c} \text{Hg} \\ 1.66 \pm 0.3 \\ 1.21 \pm 0.42 \\ 4.34 \pm 1.35 \\ 1.43 \pm 0.37 \\ 0.41 \pm 0.23 \\ \hline 7.81 \\ 0.26 \\ 1.70 \\ 4.36 \\ 1.54 \\ 0.27 \\ \hline 0.17 \\ 0.31 \\ 0.21 \\ 1.57 \pm 0.32 \\ 0.26 \pm 0.11 \end{array}$	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2	$\begin{array}{c ccccc} & & & & & & Pb \\ \hline 3.8 & 0.14\pm0 \\ \hline 3.8 & 0.14\pm0 \\ \hline 7.5 & 24.6\pm1 \\ 2.4 & 1.52\pm0 \\ \hline 5.4 & 0.47\pm1 \\ \hline 2.5 & 3.65\pm0 \\ \hline & 1.10 \\ \hline 5 & 0.02 \\ \hline 3 & 0.02 \\ \hline 1 & 0.72 \\ \hline 3 & 0.02 \\ \hline 4 & 0.72 \\ \hline 5 & 0.02 \\ \hline 5 & 0$	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.17 5.67± 0.93 1.44±0 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72 5 0.21 5 0.21 5 0.52 8 4.4 0.23 0.53±1	$\begin{array}{c cccc} & & & & & \\ \hline 1.2 & 0.41 \pm \\ 2.0 & 0.93 \pm (\\ 2.1 & 0.34 \pm (\\ 0.2 & 1.31 \pm 0 \\ 2.1 & 0.34 \pm (\\ 0.2 & 0.26 \pm \\ 3 & 1.84 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 3 & 4.33 \\ 7 & 1.5 \\ 1 & 0.44 \\ 2 & 3.79 \\ 1.22 \\ 2.8 & 7.3 \pm 2 \\ 0.2 & 0.09 \pm 0 \end{array}$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 6 44 2 40 9 73 5 42 2.1 580±60 0.03 4.6±1.9	$\begin{array}{c} C_{\rm org},\% \\ \hline 24.2\pm2.3 \\ \rm nd \\ \rm nd \\ 22.9\pm1.4 \\ 20.3\pm1.7 \\ 28.5 \\ 26.1 \\ 27.5 \\ 27.4 \\ 25.8 \\ \rm nd \\ 22.9 \\ 21.5 \\ 21.1 \\ \rm nd \\ 0.6\pm0.1 \\ \end{array}$
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end whole soft tissues shell foot	$\begin{array}{c} Cu\\ 15.4\pm 2.6\\ 7.9\pm 2.7\\ 0.6\pm 0.2\\ 7.9\pm 3.2\\ 11.3\pm 3.5\\ 34.0\\ 24.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 4.8\\ 21.3\\ 8.6\\ 879\pm 75\\ 16.6\pm 4.8\\ 9.2\\ \end{array}$	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125	$\begin{array}{c} \text{Hg} \\ 1.66\pm0.3 \\ 1.21\pm0.42 \\ 4.34\pm1.35 \\ 1.43\pm0.37 \\ 0.41\pm0.23 \\ 7.81 \\ 0.26 \\ 1.70 \\ 4.36 \\ 1.54 \\ 0.27 \\ 0.17 \\ 0.31 \\ 0.21 \\ 1.57\pm0.32 \\ 0.26\pm0.11 \\ 0.98 \end{array}$	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 0.02 5 0.02 3.00 0.02 3.00 0.72 0.72 2.44 3.66 0.72 2.2 24.3 ± 1 2.8 1.56 ± 0 3 1.84	Sb 0.08 4.95± 11.3 8.19± 0.16 0.82±0 0.13 5.67± 0.93 1.44±0 0 6.32 5 11.3 5 13.7 0 2.09 7 1.72 5 0.74 3 0.21 5 0.52 8 4.4 0.35 0.53±1 4 0.32	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 6 44 2 40 9 73 5 42 2.1 580±60 0.03 4.6±1.9 6 120	$\begin{array}{c} C_{\rm org},\% \\ 24.2\pm2.3 \\ \rm nd \\ \rm nd \\ 22.9\pm1.4 \\ 20.3\pm1.7 \\ 28.5 \\ 26.1 \\ 27.5 \\ 27.4 \\ 25.8 \\ \rm nd \\ 22.9 \\ 21.5 \\ 21.1 \\ \rm nd \\ 0.6\pm0.1 \\ \rm nd \\ \end{array}$
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=73 mm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end tube, soft tissues shell foot	$\begin{array}{c} Cu\\ 15.4 \pm 2.6\\ 7.9 \pm 2.7\\ 0.6 \pm 0.2\\ 7.9 \pm 3.2\\ 11.3 \pm 3.5\\ 34.0\\ 24.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 4.8\\ 21.3\\ 8.6\\ 879 \pm 75\\ 16.6 \pm 4.8\\ 9.2\\ 42.5\\ \end{array}$	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284	$\begin{array}{c} \text{Hg} \\ 1.66 \pm 0.3 \\ 1.21 \pm 0.42 \\ 4.34 \pm 1.35 \\ 1.43 \pm 0.37 \\ 0.41 \pm 0.23 \\ \hline 0.41 \pm 0.23 \\ \hline 0.41 \pm 0.23 \\ \hline 0.26 \\ 1.70 \\ 4.36 \\ 1.54 \\ 0.27 \\ \hline 0.17 \\ 0.31 \\ 0.21 \\ 1.57 \pm 0.32 \\ 0.26 \pm 0.11 \\ \hline 0.98 \\ 0.17 \\ \hline \end{array}$	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 0.02 5 0.02 3.00 0.02 3.00 0.47 0.72 0.72 0.72 0.72 2.43 3.66 ± 0 3.2 24.3 ± 1 2.8 1.56 ± 0 3 1.84 1.42 1.42	Sb 0.08 $4.95\pm$ 11.3 $8.19\pm$ 0.16 $0.82\pm$ 0 0.93 $1.44\pm$ 0 0.633 0.335 5 11.35 5 13.72 0.77 0.726 3 0.215 5 0.525 8 4.44 0.35 $0.53\pm$ 4 4 0.325 2 1.34	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.1 $0.34\pm$ (2.25 $1.31\pm$ (2.1 $0.34\pm$ (2.1 $0.34\pm$ (3 1.84 3 4.33 7 1.53 9 0.33^{+} 2 1.53 4 1.66 1 0.44 2 3.79 1.22 0.94 2.8 $7.3\pm$ 2 0.2 0.56 4 1.52	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 4 44 2 40 9 73 5 42 2.1 580±60 0.03 4.6±1.9 6 120 2 3110	$\begin{array}{c} C_{\rm org}, \% \\ 24.2\pm 2.3 \\ \rm nd \\ \rm nd \\ 22.9\pm 1.4 \\ 20.3\pm 1.7 \\ 28.5 \\ 26.1 \\ 27.5 \\ 27.4 \\ 25.8 \\ \rm nd \\ 22.9 \\ 21.5 \\ 21.1 \\ \rm nd \\ 0.6\pm 0.1 \\ \rm nd \\ 23.6 \end{array}$
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=73 mm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end tube, soft tissues shell foot gills mantle	$\begin{array}{c} Cu\\ 15.4 \pm 2.6\\ 7.9 \pm 2.7\\ 0.6 \pm 0.2\\ 7.9 \pm 3.2\\ 11.3 \pm 3.5\\ 34.0\\ 24.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 42.8\\ 8.7\\ 4.6\\ 26.3\\ 4.8\\ 21.3\\ 8.6\\ 879 \pm 75\\ 16.6 \pm 4.8\\ 9.2\\ 42.5\\ 45.5\\ \end{array}$	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452	$\begin{array}{c} \text{Hg} \\ 1.66\pm0.3 \\ 1.21\pm0.42 \\ 4.34\pm1.35 \\ 1.43\pm0.37 \\ 0.41\pm0.23 \\ 7.81 \\ 0.26 \\ 1.70 \\ 4.36 \\ 1.54 \\ 0.27 \\ 0.17 \\ 0.31 \\ 0.21 \\ 1.57\pm0.32 \\ 0.26\pm0.11 \\ 0.98 \\ 0.17 \\ 0.44 \\ \end{array}$	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 0.02 3.00 0.02 3.00 0.04 0.72 0.72 2.43 ± 1 2.43 ± 1 2.8 1.56 ± 0 3.12 24.3 ± 1 2.8 1.56 ± 0 3.12 24.3 ± 1 2.8 1.56 ± 0 3.156 ± 0 3.42 3.42 3.44	Sb 0.08 $4.95\pm$ 11.3 $8.19\pm$ 0.16 $0.82\pm$ 0 0.15 11.3 5 11.3 5 13.7 0 2.09 7 1.72 5 0.74 3 0.21 5 0.52 8 4.4 0.2 $13.1\pm$ 0.35 $0.53\pm$ 4 0.32 2 1.34 5.4 5.4	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.1 $0.34\pm$ (0.2 $0.26\pm$ 3 1.84 3 4.33 7 1.53 0 0.3^{2} 1 0.42 2 1.53 4 1.64 1 0.44 2 3.79 1.22 $0.09\pm$ (2.8 7.3 ± 2 0.2 0.54 4 1.65 0.2 0.54	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 175 7 82 5 173 4 44 2 40 9 73 5 42 2.1 580±60 0.03 4.6±1.9 5 120 2 3110 4 560	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=88 mm	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end tube, soft tissues shell foot gills mantle	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8 9.2 42.5 45.5 8.5	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452 191	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32 0.26±0.11 0.98 0.17 0.44 0.08	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5 56.8	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 1.10 5 0.02 3.00 0.02 3.00 0.47 0.72 0.72 0.72 0.72 0.72 0.72 0.72 0.43 0.2 24.3 ± 1 2.8 1.56 ± 0 3 1.84 1.42 5 5 4.1 3 2.72	$\begin{array}{c ccccc} & & & & & & & \\ \hline 0.08 & 4.95 \pm \\ 11.3 & 8.19 \pm \\ 0.16 & 0.82 \pm 0 \\ 0.1 & 5.67 \pm \\ 0.93 & 1.44 \pm 0 \\ 0 & 6.33 \\ 5 & 11.3 \\ 5 & 13.7 \\ 0 & 2.09 \\ 7 & 1.72 \\ 5 & 0.74 \\ 3 & 0.21 \\ 5 & 0.52 \\ 8 & 4.4 \\ 10.2 & 13.1 \pm \\ 0.35 & 0.53 \pm 1 \\ 4 & 0.32 \\ 2 & 1.34 \\ 5.4 \\ 2 & 0.00 \\ \end{array}$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=73 mm Archivesica gigas (n=1), shell length=88 mm Phelliactis pabista (n=1)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end tube, soft tissues shell foot gills mantle shell whole	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8 9.2 42.5 45.5 8.5 69.7	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452 191 8800	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32 0.26±0.11 0.98 0.17 0.44 0.08 0.23	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5 56.8 172.1	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 0.02 5.4 $0.47\pm$ 5.4 $0.47\pm$ 5.4 0.02 3.00 0.02 3.00 0.47 0.72 0.72 0.72 0.72 2.43 ± 1 2.443 ± 1 2.8 1.56 ± 0 3.156 ± 0 3.84 1.42 1.56 ± 0 3.2 24.3 ± 1 3.2 24.3 ± 1 3.2 24.3 ± 1 3.45 ± 0 1.84 3.42 3.42 $5.4.12$ 1.72 3.2 2.72 1.72 7.72	$\begin{array}{c ccccc} & & & & & & & \\ \hline 0.08 & 4.95\pm\\ 11.3 & 8.19\pm\\ 0.16 & 0.82\pm0\\ 0.1 & 5.67\pm\\ 0.93 & 1.44\pm0\\ 0 & 6.32\\ 5 & 11.3\\ 5 & 13.7\\ 0 & 2.09\\ 7 & 1.72\\ 5 & 0.74\\ 3 & 0.22\\ 7 & 1.72\\ 5 & 0.74\\ 3 & 0.21\\ 5 & 0.52\\ 8 & 4.4\\ 10.2 & 13.1\pm\\ 0.35 & 0.53\pm1\\ 4 & 0.32\\ 2 & 1.34\\ 5.4\\ 2 & 0.00\\ \hline 9 & 1.82\\ \hline \end{array}$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Zn 0.1 255±27 0.23 231±27 0.25 410±46 0.13 105±12 0.1 112±13 4 101 5 380 7 82 5 173 4 40 9 73 5 42 2.1 580±60 0.03 4.6±1.9 5 120 2 3110 4 560 6 20 1 3317	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd nd 23.6 nd nd
Taxon (number of specimens)Riftia pachyptila (n=3), tube length= 10-21.5 cmRiftia pachyptila (n=1), tube length= 51 cmRiftia pachyptila (n=1), tube length= 115 cmNuculana grasslei (n=15)Archivesica gigas (n=1), shell length=73 mmArchivesica gigas (n=1), shell length=88 mmPhelliactis pabista (n=1)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end tube, soft tissues shell foot gills mantle shell whole	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8 9.2 42.5 45.5 8.5 8.5 69.7 51.7	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452 191 8800 51	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32 0.26±0.11 0.98 0.17 0.44 0.08 0.23 0.18	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5 56.8 172.1 230.0	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 0.02 3.00 0.47 0.72 0.72 2.43 ± 1 2.8 1.56 ± 0 3 1.84 1.22 24.3 ± 1 2.8 1.56 ± 0 3 1.84 5 4.1 3 2.72 1 70.5 0 0.17	$\begin{array}{c ccccc} & & & & & & & \\ \hline 0.08 & 4.95\pm\\ 11.3 & 8.19\pm\\ 0.16 & 0.82\pm0\\ 0.1 & 5.67\pm\\ 0.93 & 1.44\pm0\\ 0 & 6.32\\ 5 & 11.3\\ 5 & 13.7\\ 0 & 2.09\\ 7 & 1.72\\ 5 & 0.74\\ 3 & 0.22\\ 5 & 0.52\\ 8 & 4.4\\ 10.2 & 13.1\pm\\ 0.35 & 0.53\pm1\\ 4 & 0.32\\ 2 & 1.34\\ 2 & 1.34\\ 2 & 0.00\\ \hline 0 & 1.82\\ 7 & 0.26\\ \hline \end{array}$	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.1 $0.34\pm$ (0.2 $0.26\pm$ 3 1.83 4 3.6 4 1.5 0 0.3^{2} 1.53 1.64 1 0.44 2 3.79 1.22 $0.09\pm$ (2.8 $7.3\pm$ 2 0.2 0.54 2 0.54 3 1.8 9 0.14 3 1.8 5 0.12	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd nd nd 23.6 nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=73 mm Archivesica gigas (n=1), shell length=88 mm Phelliactis pabista (n=1) Munidopsis alvisca (n=1)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, anterior end whole soft tissues shell foot gills mantle shell whole chitin spawn	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8 9.2 42.5 45.5 8.5 69.7 51.7 39.9	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452 191 8800 51 26	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32 0.26±0.11 0.98 0.17 0.44 0.08 0.23 0.18 0.11	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 2.9 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5 56.8 172.1 230.0 17.2	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 1.10 1.52 ± 0 5.4 $0.47\pm$ 2.5 3.65 ± 0 3.00 0.02 3.00 $0.47\pm$ 0.72 2.43 ± 1 2.8 1.56 ± 0 3.2 24.3 ± 1 2.8 1.56 ± 0 3 1.82 5 4.1 3 2.72 1 70.9 0 0.17 2 3.58	Sb 0.08 $4.95\pm$ 11.3 $8.19\pm$ 0.16 $0.82\pm$ (0 0.11 $5.67\pm$ 0.93 $1.44\pm$ 0 6.32 5 11.3 5 11.5 5 13.7 0 2.09 7 1.72 5 0.52 8 4.4 0.25 $0.52\pm$ 4 0.32 2 $1.34\pm$ 2 1.34 2 1.34 2 0.00 9 1.82 7 0.26 7 0.26	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.1 $0.34\pm$ (0.2 $0.26\pm$ 3 1.83 4 3.6 7 1.52 0 0.3^{2} 1 0.42 2 1.53 4 1.66 1 0.42 2 3.79 1.22 $0.09\pm$ (2 0.52 0.2 $0.09\pm$ (2 0.59 4 1.6 1 0.42 2 0.59 4 1.52 0.2 $0.09\pm$ (2 0.54 9 0.116 3 0.63	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd nd nd nd nd nd
Taxon (number of specimens) Riftia pachyptila (n=3), tube length= 10-21.5 cm Riftia pachyptila (n=1), tube length= 51 cm Riftia pachyptila (n=1), tube length= 115 cm Nuculana grasslei (n=15) Archivesica gigas (n=1), shell length=73 mm Archivesica gigas (n=1), shell length=88 mm Phelliactis pabista (n=1) Munidopsis alvisca (n=1)	Tissue trophosome obturaculae opisthosome vestimentum tube trophosome obturaculae opisthosome obturaculae opisthosome vestimentum gonades tube tube, posterior end tube, middle part tube, anterior end tube, middle part tube, anterior end tube, soft tissues shell foot gills mantle shell whole chitin spawn remaining	Cu 15.4±2.6 7.9±2.7 0.6±0.2 7.9±3.2 11.3±3.5 34.0 24.3 42.8 8.7 4.6 26.3 4.8 21.3 8.6 879±75 16.6±4.8 9.2 42.5 45.5 8.5 69.7 51.7 39.9 122	Fe 785±80 92±8 nd 10±3 102±14 463 451 131 110 62 112 nd 215 92 8905±765 101±23 125 284 452 191 8800 51 26 107	Hg 1.66±0.3 1.21±0.42 4.34±1.35 1.43±0.37 0.41±0.23 7.81 0.26 1.70 4.36 1.54 0.27 0.17 0.31 0.21 1.57±0.32 0.26±0.11 0.98 0.17 0.44 0.08 0.23 0.18 0.11 0.25	Mn 10.2±3 19.3±7 10.1±2 15.5±5 12.2±2 7.8 17.6 13.8 8.6 6.1 6.2 9.5 5.5 6.8±3 5.25±2 10.8 8.6 11.5 56.8 172.1 230.0 17.2 367.5	Pb 3.8 0.14 ± 0 7.5 24.6 ± 1 2.4 1.52 ± 0 5.4 0.47 ± 1 2.5 3.65 ± 0 1.10 0.02 3.00 0.02 3.00 0.04 0.72 0.72 2.43 3.66 ± 0 3.22 24.3 ± 1 2.8 1.56 ± 0 3 1.84 1.42 5 4.11 3 $3.2.72$ $1.70.9$ 0 0.17 2 3.58 5 7.06	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Se 1.2 $0.41\pm$ 2.0 $0.93\pm$ (2.1 $0.34\pm$ (2.25 $1.31\pm$ (2.1 $0.34\pm$ (2.1 $0.34\pm$ (3 1.83 3 4.33 7 1.52 9 0.32 2 1.53 4 1.60 1 0.42 2 3.79 1.2 $0.09\pm$ (2 0.52 0.2 $0.09\pm$ (2 $0.59\pm$ (4 1.52 0.2 $0.09\pm$ (2 $0.59\pm$ (4 1.52 0.52 $0.09\pm$ (3 0.63 5 0.10^{-10}	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	C _{org} , % 24.2±2.3 nd nd 22.9±1.4 20.3±1.7 28.5 26.1 27.5 27.4 25.8 nd 22.9 21.5 21.1 nd 0.6±0.1 nd 23.6 nd nd nd nd nd nd nd nd nd nd nd nd 23.6

Table 3. Content (mg kg⁻¹ dry weight) of some elements in organism tissues and whole organisms inhabited the Southern trough of the Guyamas Basin

n.d.- not determined (not analyzed)





Figure 2. Metal contents (mg kg⁻¹ dry weight) in the different organs of vestimentifera *Riftia pachyptila* at the Guaymas Basin's vent field: a) As, Ba, Cu, Fe, Mn and Zn; b) Ag, Cd, Co, Cr, Hg, Pb, Sb and Se.

order of magnitude higher) than in other tissues (Table 3, Figures 3a, 3b).

The metal distribution in the soft tissues of the second dominant bivalve mollusk *Nuculana grasslei* is remarkable for having very high or the highest contents of many elements, including: Fe (8904 mg kg⁻¹), Ba (1440 mg kg⁻¹), Cu (873 mg kg⁻¹), Zn (580 mg kg⁻¹), Ag (64 mg kg⁻¹), As (56.3 mg kg⁻¹), Pb (24.3 mg kg⁻¹), Sb (13.3 mg kg⁻¹) and Se (7.3 mg kg⁻¹). As a rule, the difference between metal contents (except Mn) in the soft body compared to the shell was significant, reaching up to two orders of magnitude.

In the whole body of the *Phelliactis pabista* specimens, which were collected from areas impacted by hot shimmering water, elevated contents of Cd (5.7 mg kg⁻¹), Cu (70 mg kg⁻¹), Fe (8800 mg kg⁻¹), Mn (172 mg kg⁻¹), Pb (71 mg kg⁻¹), and Zn (3317 mg kg⁻¹) were found. *Phelliactis pabista* were collected in the microhabitats of the vesicomyid clam *Archivesica gigas*. Many specimens were attached to the shells of these clams.

In the chitin and remaining tissues (gills) of the *Munidopsis alvisca* crab, elevated concentrations of Cu (122 mg kg⁻¹) and Mn (367 mg kg⁻¹) were detected. These levels were higher than those observed in the organs of *Munidopsis alvisca* prey *Riftia* and *Archivesica gigas*.

In the whole body of *Spongia* specimens, collected where hot fluids were emitted, maximum Mn content (2915 mg kg⁻¹) was detected, approximately two orders of magnitude higher than in the other organisms. The other chemical elements studied also demonstrate rather high concentrations in *Spongia* (Table 3). Sponges are filterfeeding organisms that utilize organic matter and bacterial biomass as well as mineral suspensions. Their biochemical functioning and siliceous mineral skeletons lead to the formation of biostructures with very high porosity, which contributes to their effective adsorption and absorption capabilities.

4. Discussion

The geochemical environmental conditions of the Southern trough of the Guaymas Basin apparently result in some peculiarities in fluid composition. In particular, Mn concentration is distinctly higher than Fe concentration in hot fluids of the first type compared to warm fluids of the second type. This differs from the hydrothermal vent fluids of the Mid-Atlantic Ridge and 9° 50' N of the East Pacific Rise, where Fe is found in higher concentrations than Mn (Douville et al., 2002; Von Damm, 2000). Magnesium is completely absent in pure end-member hydrothermal fluids due to its uptake by basalts during hydrothermal circulation (Von Damm, 1990). The relatively high Mg concentrations in our water samples are evidence of a high degree of fluid dilution with the seawater. On the other hand, the levels of trace metals in these fluids are much lower than in the end-member fluids of the Guaymas hydrothermal vent field (Von Damm *et al.*, 1985), which also implies that fluids are strongly diluted by seawater during their mixing. We also cannot exclude partial precipitation of particles from the fluids inside a sedimentary cover during neutralization and cooling. The Fe/Mn molar ratios are 5.0 and 0.47 for fluids of the first and second types respectively, with an average Fe/Mn ratio of 2.7. This is much lower than the basalt Fe/Mn ratio (50-60), but rather close to the Fe/Mn of 3 for metalliferrous sediments (Dymond *et al.*, 1973). This suggests that, due to various biogeochemical processes within the sedimentary cover, the behavior of Fe and Mn in fluids ascending through the sediment may be altered, resulting in enrichment of Fe relative to Mn in the first type fluids or vice versa in the second type.

Since hydrothermal vestimentiferas *Riftia* was first discovered only about 30 years ago, some features of its metabolism are not fully elucidated. The trophosome is known to host intracellular symbiotic bacteria, performing sulfide oxidation and carbon fixation. Based on this, the high C_{org} content (28.53 %) detected in trophosome is quite natural and can be attributed to accumulation of endosymbiotic bacteria. A rather small difference (about 8 %) between the highest and lowest C_{org} concentrations in the Riftia organs might be attributed, in our opinion, to the presence of bacterial biomass in the form of microbial mats and overgrowth on the *Riftia* tubes. The vestimentiferan growth rate is known to range from 10 (Fustec et al., 1988) to 50 cm per year (Tunnicliffe, 1991). Thus, it follows that our examined Riftia individuals are rather young - between one (tube length 10-21 cm) and five (tube length 51 cm) years old. Our data do not allow us to examine differences in the trace metal contents of different organs in these two groups; however, data from Ruelas-Inzunza et al. (2005) seem to show that only concentrations of Cd and Fe in vestimentum increase with the size of specimens. The rest of the metals did not show any relationship to length. Ruelas-Inzunza et al. (2005) also found that the trophosome is an organ with high accumulation of Co, Cu and Fe. Our data support this conclusion for Cu and Fe. The calculation of average metal contents in two different-sized groups of *Riftia* shows that the trophosome is a target organ for Fe (624 mg kg⁻¹), Cu (24.7 mg kg⁻¹), Ag (2.19 mg kg⁻¹) and Hg (4.73 mg kg⁻¹). It should be noted that our data on the distribution of the majority of metals in different Riftia organs are similar in orders of magnitude to earlier published findings for the Guaymas Basin, with the exception of Cu in the opistosome, Fe in the vestimentum and Hg in the trophosome and vestimentum; in these cases our data are approximately one order of magnitude lower (Table 4).

Using data from Tables 3 and 5 (Demina *et al.*, 2007), inter-site comparisons of metal content can be conducted for organisms taken from geochemically different vent fields: the 9° 50' N EPR and the Guaymas Basin. In *Riftia* organs the Mn contents are about 3 times higher in the trophosome and obturaculum and 24 times higher in the tubes of the Guaymas Basin's specimens relative to specimens taken





Figure 3. Distribution of metals in organs of vesicomyid clam Archivesica gigas: a) Ba, Cu, Fe, Mn and Zn; b) Ag, As, Au, Cd, Co, Cr, Hg, Pb, Sb and Se.

1											
Organ	Ba	Cd	Со	Cr	Cu	Fe	Hg	Mn	Pb	Zn	Reference
Vestimetntifera	a Riftia pa	achyptila									
trophosome	98	1.9	2.8	2.9	13.8	493	n.d.	0.7	9	428	Lein et al., 1989
trophosome	n.d.	1.7	2.2	3.4	26.4	521	22.2	4.1	1.9	245	Ruelas-Inzunza et al., 2005
vestimentum	74	1.5	0.1	7	27	4900	n.d.	31	<5	250	Lukashin et al., 1990
vestimentum	n.d.	3.7	0.7	3.4	11.6	297	22.5	6	2	599	Ruelas-Inzunza et al., 2005
opistosome	n.d.	5	n.d.	2.1	117	333	n.d.	24	n.d.	1613	Lein et al., 1989
tube	50	1	1.5	<2	16.7	420	n.d.	30.7	5.4	195	Lukashin et al., 1990
Vent clam Arch	hivesica g	rigas									
gills	230	11	1.5	8.3	12	400	n.d.	6.6	5	1000	Lein et al., 1989
whole soft tissues	350	9.5	5	2.5	18	290	n.d.	11	5	200	Lukashin et al., 1990
shells	n.d.	10	n.d.	6	10	54	n.d.	56	22	135	Lukashin et al., 1990
Vent clam Vesi	comya gi	gas									
gills	n.d.	115.2	n.d.	n.d.	8.26	403	4.96	18	2.89	845	Puolos Ingunzo et al. 2002
mantle	n.d.	12.3	n.d.	n.d.	29.7	277	1.1	10.6	3.67	419	Rucias-inzunza el al., 2003

Table 4. Levels of some trace elements (mg kg⁻¹ dry weight) in the invertebrates inhabited the Guaymas Basin's hydrothermal vent field, based on published data

n.d. is not determined (not analyzed)

Table 5. Average content	* of some elements in organ	s of mussels, clams and	d vestimentiferas from	the 9°50' degree	N hydrothermal field (Demina et al.,
2007)						

Taxon	Organ	Ag	As	Ва	Cd	Co	Cr	Cu	Fe	Hg	Mn	Pb	Zn	C _{org} , %
	ailla	15.44	7.50	46	4.66	42.75	5.91	166.4	214.4	0.18	4.32	0.40	120.1	27.1
	gins	1.64	3.22	28	0.15	30.76	5.35	6.1	2.6	0.04	1.67	0.45	15.1	5.7
	mantla	4.58	2.23	70	0.30	5.13	3.15	18.6	25.1	0.08	1.18	0.87	41.9	25.5
	mantie	2.82	2.31	15	0.39	4.49	1.07	1.2	7.7	0.04	0.11	1.15	0.2	6.8
Dathum o di olug thoma of lug	faat	2.00	3.15	563	0.07	6.14	32.39	5.8	36.8	0.19	2.37	1.51	30.2	23.3
Bainymoalolus inermojilus	s 100t	0.09	3.73	583	0.08	3.27	5.20	2.3	35.0	0.20	0.43	0.78	1.6	3.6
	romaining	4.68	2.37	30	0.85	6.08	1.27	27.9	101.3	0.14	1.12	2.36	65.6	29.2
	remaining	0.10	0.21	18	0.84	2.06	0.72	13.4	32.0	0.12	0.11	3.28	18.7	6.3
	ahall	0.02	1.98	67	0.21	2.44	0.40	7.6	52.1	0.05	4.41	6.98	8.2	2.4
	snell	0.00	2.16	62	0.26	0.69	0.19	4.5	2.2	0.01	2.07	3.74	5.9	1.6
	gills	2.28	9.43	94	1.55	3.16	74.0	66.8	377.9	0.05	3.78	1.73	260.2	29.5
	mantle	3.79	3.79	34	2.06	3.63	10.8	32.9	170.0	0.06	49.22	8.36	320	23.8
Archivesica gigas	foot	4.64	4.64	30	0.12	3.42	1.54	18.3	140.0	0.07	2.53	102.1	400	28.3
Bathymodiolus thermofilu Archivesica gigas Riftia pachyptila	remaining	7.27	3.61	23	1.1	2.45	3.19	21.4	100.8	0.04	4.17	1.61	145.1	26.0
	shell	0.03	1.03	42	0.01	2.8	1.32	4.68	24.8	0.05	1.94	8.63	12	0.3
	tranhagama	3.83	5.82	39	0.23	17.05	1.09	23.3	389.3	0.51	3.19	11.95	157.5	28.9
	uophosome	4.13	1.87	15	0.19	13.48	0.70	23.8	171.0	0.22	2.45	17.71	97.9	5.1
	- 1- 4	1.52	2.13		0.72	9.42	4.62	18.9	1035	0.14	13.06	8.44	355.3	24.8
	obturaculum	1.16	0.88		0.62	8.93	6.15	8.3	760.1	0.13	7.53	11.83	129.7	4.3
Diffia nachuntila	anistacama	3.67	2.96	608	0.10	3.21	4.02	20.0	232.1	0.74	166.37	7.51	166.4	32.7
кіјна распурша	opistosome	2.75	2.60	796	0.01	1.41	1.83	19.8	14.2	0.11	63.88	5.03	63.9	0.0
		2.22	1.29	75	0.25	13.28	2.03	13.1	164.0	0.51	31.32	2.71	72.0	24.5
	vesumentum	1.87	0.38	49	0.25	8.49	1.22	12.5	48.9	0.37	38.82	0.55	22.0	1.0
	4.h.	0.56	2.12	50	0.23	4.01	2.04	18.9	118.0	0.29	0.31	3.47	26.1	23.6
	tube	0.20	0.79	14	0.27	0.49	2.15	10.1	77.5	0.19	0.02	5.45	9.4	1.6

* metal content - in mg kg $^{-1}$ dry weight; for each organ the first line is average content, the second one is \pm standard deviation.

from the 9°50' N EPR. A similar excess in Guaymas Basin's specimens was found for As (2 to 12 times higher), Cd (4 to 18 times higher), and Hg (up to 9 times higher). Meanwhile Cu, Fe and Zn showed similar levels in both cases, but the Ag, Co and Pb contents were several times lower in the Guaymas Basin's *Riftia* specimens. A rather high standard deviation of average contents of many metals obviously testifies the lack of significant difference in metal contents in specimens of these two fields.

A comparison of metal levels in bivalves showed that only in the external organs (gills and shells) there is a noticeable accumulation (an order of magnitude higher) of essential metals (Fe, Mn and Zn) in the Guaymas Basin's specimens. Hg contents in the soft tissues are 10 times higher in Guaymas Basin's specimens than in specimens collected from the 9° 50' N EPR. Contents of these metals are at similar levels in both cases for other organs. Specimens from the 9° 50' N EPR contain only three elements (As in gills, Ag in foot and Co in mantle, foot and shell) at about 10 times higher than the level of Guaymas Basin's specimens.

Thus, we cannot conclude that levels of the majority of metals in organism tissues are a reflection of their levels in the fluids, since majority of metals show much higher concentrations in the fluids of the 9° 50' N EPR hydrothermal region in comparison to the Guaymas Basin (Von Damm, 2000; Von Damm *et al.*, 1985).

The gills of the vesicomyid clams are known to contain endosymbiotic bacteria. Along with symbiotrophy, a nutritional strategy of this clam may include suspensionfiltering feeding. High amounts of Cd, Cu, Fe, Mn, Pb, S, and Zn were detected in the whole soft tissues (where gills have a larger contribution of biomass compared to other organs) of a similar vesicomyid clam, Archivesica gigas, from 21° N at the East Pacific Rise (Roesijadi and Crecelius, 1984). The data of Ruelas-Inzunza et al. (2003) on metal distribution in different tissues of the vent clam Vesicomya gigas from the Guaymas Basin indicate that the highest amounts of Cd, Fe, Hg, Mn and Zn were detected in the gills, while highest contents of Cu and Pb occurred mainly in the mantle. The difference between Cd concentrations in the gills and mantle did not exceed one order of magnitude, while for the rest of the metals it was only 1.5-4 times. Slightly greater amounts of metal accumulation were shown in the gills of Archivesica gigas, relative to other soft tissues (Lein et al., 1989; Lukashin et al., 1990). According to our data, Ag, As, Au, Cd, Cu, Fe, Pb, and Sb are accumulated in mantle to a greater extent than in the gills of the vesicomyid clam Archivesica gigas (Figure 3). This might be reasonably explained by a distinguishing feature of the Guaymas Basin's environmental conditions: a significant amount of production in the bacterial mats, covering sulfide ores (27 mg C_{org} m⁻²day⁻¹, Lein et al., 1988) and contained in the upper 50 cm sedimentary layer (91 mg C_{org} m⁻²day⁻¹, Gal'chenko et al., 1989) that is several times higher than in the other hydrothermal fields. We may suppose that in such an environment, which is highly enriched in organic carbon, the clam population may receive nutrients by suspensionfeeding to a greater extent than is observed as a result of symbiotrophy. In this case, in a mantle where chemical elements are assimilated mainly from food rather than from water via gills, metals could be highly accumulated relative to gills.

The shells of the vesicomyid clam Archivesica gigas are an important target for Ba, Mn, and to a lesser extent of Fe (Figure 3a), however for other metals, they play only a small role. Taking into account the large mass of the shell relative to the soft tissue of clams (in which the former may reach one order of magnitude higher that the latter), we can suggest that shells, which have accumulated trace metals during biomineralization and adsorption, might serve as a great reservoir for many metals. The second abundant bivalve mollusk Nuculana grasslei lives on substratum saturated with hydrocarbons (Allen, 1993). Unlike similar species, this animal has an extremely thick periostratum (an exterior part of the shell) that is considered to be an adaptation to functioning in an acidic environment enriched in sulfides. Its nutritional source has not yet been studied completely, but some researchers regard Nuculana grasslei as a symbiotrophic organism containing bacteria in its gills that can combine symbiotrophy with filter-feeding. This might lead to a significant bioaccumulation of a majority of metals in the whole body of Nuculana grasslei.

The high levels of some essential metals (Cu and Mn) in the soft tissues of the Munidopsis alvisca crab seem to be caused by metabolic peculiarities as well as type of feeding, namely, preying on other organisms such as the symbiotrophic Riftia pachyptila and the clams Archivesica gigas and Nuculana grasslei. Phelliactis pabista, which are typically predators, can scavenge both fragments of bacterial mats and organic matter into the vent fields and thus might enhance their metal bioaccumulation. It is important to note that Kádár et al. (2007) revealed trophic level-specific variations in essential metal accumulation in the Menez Gwen hydrothermal community of the Mid Atlantic Ridge, showing a general trend of biomagnification of Cu, Fe, and Zn from primary producers (endosymbiont bacteria) to primary (symbiont reliant species and filter-feeders) and secondary consumers (predators and scavengers).

The considerable Mn accumulation (much higher than other metals) in *Spongia* might be caused by the predominance of Mn in the water column, influenced by fluids of the first type, compared to other metals. In the body of *Spongia*, the Fe/Mn ratio is 0.24, on the same order of magnitude as fluids of the first type (0.5).

As one can see from Table 3, Mn content in the majority of dominant organisms studied is considerably lower than Fe content (from one to two orders of magnitude). In the different tissues of the dominant animals vestimentifera *Riftia*, vesicomyidae clam *Archivesica gigas* and bivalve mollusk *Nuculana*, the Fe/Mn ratio varies from 0.8 to 1309. The lower values (≤ 20) were detected in exterior organs such as the tubes of *Riftia* and the shells of *Archivesica* gigas. The higher values were found in the interior organs, especially those linked with endosymbiotic bacteria, which includes the trophosome of Riftia and the gills of the clam Archivesica gigas. The highest Fe/Mn ratio was found in the soft tissues of the bivalve mollusk Nuculana. It should be mentioned that Fe/Mn ratios in our fluid samples vary from 0.47 to 5.0. This differentiation in Fe/Mn ratios seemingly reflects the different forms (or species) of Fe and Mn, which determine their bioavailability. Due to its slower oxidationreduction kinetics. Mn is able to exist in the dissolved form of Mn (+2) longer than Fe, where it is utilized by bacteria and involved in relatively rapid microbially catalyzed Mn oxidation followed by formation of bacterial aggregates that are not commonly assimilated by benthic organisms (Campbell *et al.*, 1988). Bacterial aggregates are taken up by zooplankton, followed by excretion of the Mn-enriched fecal pellets found in the settling material taken by sediment traps (Tambiev and Demina, 1992). Possibly, this process may lead to additional enrichment in Mn of the surface sediments, which in turn provides the benthic flux of Mn.

Studies of the trace metal levels of marine mollusks from areas of the Gulf of California influenced by mining, agriculture and urban activities have shown that areas that are apparently pristine or have few anthropogenic activities have a higher content of metals such as Cu, Fe and Pb in soft tissues (Table 6) (Cadena-Cárdenas et al., 2008). Comparison of these data with levels of metal content in the soft tissues of Nuculana grasslei presented in this paper allows us to conclude that Pb and Hg levels in mollusks from the Guaymas Basin are almost one order of magnitude higher than the levels observed in mollusks from polluted areas of the Gulf of California, whereas Fe, Zn, and Cu did not reveal any significant differences. Mn contents in mollusks from both contaminated and pristine areas are essentially higher than Mn contents in vent clams. In our opinion, this suggests the importance of the geochemical features of the Guaymas Basin, namely, the low fraction of bioavailable Mn in the water column, which leads to relatively low Mn content in the tissues of organisms despite the very high concentration of total dissolved Mn in the microhabitat. The latter can be confirmed by the low concentration factor F_c (ratio of metal content in an organism to its concentration in the surrounding water) of Mn in the whole body of symbiotrophic animals: F_c is only 5 and 66 in Riftia pachyptila and Nuculana grasslei respectively (Table 7). On the contrary, F_c of Mn in the filter-feeding and preying organisms reaches up to 2.103. For the rest of metals F₂ varies within three orders of magnitude - from 30 (Co) to $3 \cdot 10^4$ (Cd). This apparently testifies a selectivity of trace element bioaccumulation by the organisms which is determined by metal bioavailability independently of metal concentration in the water column, as well as by the feeding type of taxon. Values of F_c for different organisms of the vent community of the Menez Gwen hydrothermal field were from $7 \cdot 10^3$ to $8 \cdot 10^4$ for Fe, from 10^3 to $3 \cdot 10^4$ for Cu, and from 5•103 to 2•105 for Zn (Kádár et al., 2007), being similar to our data (Table 7).

5. Conclusions

The bioaccumulation of heavy metals in the Guaymas Basin hydrothermal vent fields occurs under very specific biogeochemical conditions associated with thick sedimentary cover, high biological productivity and high bacterial production of organic carbon. In such conditions, the target organs of many metals are not only organs containing endosymbiotic bacteria (the trophosome of vestimentifera *Riftia* or the gills of vesicomyid clam *Archivesica*) but also other organs of these organisms (such as the obturaculae and opisthosome of *Riftia* and the mantle of *Archivesica*) and even other taxonomic groups of invertebrates (Phelliactis pabista, Sponge, Munidopsis alvisca) that do not bear endosymbiotic bacteria. Possibly due to a combination of symbiotrophy with suspension-feeding, the body of the bivalve mollusk Nuculana grasslei is highly enriched relative to the shell in all the metals studied. While the lowest contents for a majority of metals were found in the shells of clams, however the masses of the shells as a rule are much higher than those of the soft tissues, and for the mentioned reason we suggest that shells might serve as a great reservoir for many metals.

Based on the variability of Fe/Mn molar ratios in diluted fluids and different tissues of organisms from the hydrothermal vent areas, we can assume that Fe and Mn are subjected to a fractionation during bioaccumulation processes. The apparent reason for this behavior might be the different chemical speciation of Fe and Mn, which determine the low bioavailability of Mn for symbiotrophic organisms. There were no significant differences between the Cd, Cu, Fe, Hg, Pb, and Zn levels in the Guaymas Basin vent clams versus the bivalve mollusks from polluted areas of the Gulf of California, which suggests the importance of the heavy metal bioavailable fraction rather than their total content in the bioaccumulation.

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Metal	Range	Bivalve species	Area	Reference		
	2231 - 2311	clams Chione subrugosa and Tellina sp.	Pabellón-Atlata wetland system in Sinaloa	Páez-Osuna et al., 1993		
	150 - 700	mussel M.strigata	Estero de Urias, Sinaloa	Ruelas-Inzunza and Páez-Osuna, 2000		
	85.6 - 397	clams C.gnidia and L.elatum	Guaymas Bay, Sonora (contaminated area)	Méndez et al., 2002		
Fe	323 - 438 154 - 558	clam Megapitaria squalida	near the phosphorite mine La Paz pristine (far from this mine)	Méndez et al., 2006		
	189 - 572	clams Ch. californiensis, M.squalida, L.elatum	different localities of the Gulf of Cali- fornia	Cadena-Cárdenas et al., 2008		
	150 - 369	mussel M.edulis				
	25 - 1247	clam Chione californiensis				
	64 - 1218	clam Chione subrugosa	the southern Gulf of California	Páez-Osuna et al., 1993		
	64 - 1944	clam <i>Tellina</i> sp.				
7	200 (mean)	mussel M.strigata	the southern Gulf of California	Ruelas-Inzunza and Páez-Osuna, 2000		
ZII	92.4 - 246	clams C.gnidia and L.elatum	Guaymas Bay, Sonora (contaminated area)	Méndes et al., 2002		
	up to 64,6	clam Megapitaria squalida	La Paz Bay (pristine area)	Méndes et al., 2006		
	49.8 - 91.8	clams Ch. californiensis, M.squalida, L.elatum	different localities of the Gulf of Cali- fornia	Cadena-Cárdenas et al., 2008		
	47.9 - 94.3	mussel M.edulis				
	4 - 5	mollusks	pristine areas along the Gulf of California			
	up to 82	forniensis Chione subrugosa and Tellina sp.	some areas along the Gulf of California	Páez-Osuna et al., 1993		
	13.4 (mean)	clams	Colorado River delta	García-Hernández et al., 2001		
Cu	4.78 - 23.04	clams C.gnidia and L.elatum	Guaymas Bay (contaminated area)	Méndez et al., 2002		
Cu	up to 8	clam Megapitaria squalida	La Paz Bay (pristine area)	Méndez et al., 2006		
	3.9 - 181	clams Ch. californiensis, M.squalida, L.elatum	different localities of the Gulf of Cali-	Cadena-Cárdenas <i>et al.</i> 2008		
	6.1-49.6	mussel M.edulis	fornia			
	0.51 - 4.03	clams C.gnidia and L.elatum	Guaymas Bay, Sonora (contaminated area)	Méndez et al., 2002		
	up to 7.8	clam Megapitaria squalida	La Paz Bay (pristine area)	Méndez et al., 2006		
Pb	3,1 (mean)	mussel Mytella strigata	south-eastern Gulf of California, Urias estuary	Soto-Jiménez et al., 2008		
	<0.03 - 9.2	clams Ch. californiensis, M.squalida, L.elatum	different localities of the Gulf of Cali- fornia	Cadena-Cárdenas et al., 2008		
	0.27 - 5.8	mussel M.edulis				
	3.8	clam Chione californiensis				
	5.5 8 7	clam Talling sp	Pabellon-Altata wetland system in Sinaloa	Paez-Osuna <i>et al.</i> , 1995		
	0.21 - 1.28	clams <i>C</i> gnidia and <i>L</i> elatum	Guavmas Bay Sonora (contaminated area)	Méndez at al. 2002		
Cd	11 1	clam Meganitaria saualida	La Paz Bay (pristine area)	Méndez et al. 2002		
	0.42 - 4.66	clams Ch. californiensis, M.squalida, L.elatum	different localities of the Gulf of Cali-	Cadena-Cárdenas et al. 2008		
	2.42 - 4.05	mussel M.edulis	fornia	Cudena Cardenas er an, 2000		
	69	clam Chione californiensis				
	80	clam Chione subrugosa	Pabellón-Altata wetland system in Sinaloa	Páez-Osuna et al., 1993		
	43	clam <i>Tellina</i> sp.				
Mn	1.59 - 26.9	clams C.gnidia and L.elatum	Guaymas Bay, Sonora (contaminated area)	Méndez et al., 2002		
	14.1 (mean)	clam Megapitaria squalida	La Paz Bay (pristine area)	Méndez et al., 2006		
	1.21 - 3.68 0 53 - 3 12	clams Ch. californiensis, M.squalida, L.elatum mussel M edulis	different localities of the Gulf of Cali- fornia	Cadena-Cárdenas et al., 2008		
Hg	0-063 (mean)	clam Chione subrugosa	south-eastern Gulf of California	Green-Ruiz et al., 2005		
0	0.23	oyster Craassostrea gigas				

Table 6. Levels of some heavy metal contents (mg kg⁻¹ dry weight) in the soft tissues of the mollusks from anthropogenically polluted and pristine areas of the Gulf of California.

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Table 7. Heavy metal content (C_w) in the whole organisms and their bioconcentration factors (F_c) at the Guaymas Basin Southern trough.

Species	Feeding type		Ag	As	Cd	Co	Cr	Cr	Cu	Fe	Mn	Pb	Sb	Zn
Vestimentifera <i>Riftia pachyptila</i>	Chemoautotrophy	C _m F _c	0.84 4•10 ³	9.9 2•10 ²	2.24 10 ⁴	0.9 82	1.8 45	1.8 45	15.6 4•10 ²	259 2•10 ²	11.7 5	5.2 6•10 ²	5.3 10 ³	163 8•10 ³
Bivalve mollusk Nuculana grasslei	Chemoautotrophy ?	C _m F	7.52 3•10 ³	7.73 4•10 ²	0.71 2•10 ³	0.4 30	2.8 10 ²	2.8 10 ²	102 1•10 ³	980 2•10 ³	5.4 66	3.83 10 ³	1.78 3•10 ²	61.8 50
Actinia Actinaria	Filter-feeding, predator	C _m F _c	-	8.2 10 ³	5.7 10 ⁴	0.3 30	1.3 50	1.3 50	69.7 4•10 ³	8800 2•10 ⁴	172 2•10 ³	71 2•10 ⁴	1.8 3•10 ²	3317 3•10 ³
Sponge Spongia	Filter-feeding	C _m F _c	0.91 4•10 ³	9.5 2•10 ²	6.64 3•10 ⁴	1 10 ²	1.7 40	1.7 40	81 2•10 ³	703 10 ³	2915 10 ³	24.7 3•10 ³	1.1 3•10 ²	200 10 ⁴

 C_m = metal content in the whole organism (mg kg⁻¹ dry weight), calculated from the concentrations of each metal in organs, constitutioning whole body, on the base of the percentage of mass of each organ.

 $F_c = C_m / C_{water}$

the zooplankton and suspended particulate matter in the biogeochemistry of the trace elements in the central region of the Gulf of California".

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